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Nano-Biosensors for Early Detection of Obesity-Related Cancer Biomarkers

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ABSTRACT

Obesity establishes a chronic, low-grade inflammatory and endocrine milieu that elevates cancer risk and worsens outcomes across multiple organs. Early interception depends on detecting composite biomarker signatures spanning inflammatory cytokines, adipokines, oncometabolites, extracellular vesicles, and tumor-derived nucleic acids at clinically actionable concentrations and in real-world matrices complicated by dyslipidemia and high protein burden. Nanoscale biosensors bring the requisite surface-to-volume ratio, quantum and plasmonic effects, and electron transport properties to transduce scarce targets with high selectivity while supporting multiplexing in small sample volumes. This review charts advances in electrochemical, field-effect transistor, plasmonic/optical, SERS, quantum dot, nanopore, and microfluidic nanotechnologies tailored to the obesity-cancer interface. It emphasizes antifouling surface chemistries that remain specific in lipid-rich plasma, aptamer and molecularly imprinted polymer recognition layers that rival antibodies, and device architectures that couple pre-concentration, sorting, and readout on a single chip. Integration with wearable microneedles and sweat or saliva sensors points toward longitudinal, minimally invasive surveillance, while machine-learning fusion of multimodal signals promises individualized risk stratification and treatment triage. Finally, we outline validation, calibration, and regulatory pathways to move nano-biosensing from elegant prototypes to robust clinical tools that detect obesity-linked cancer signals months to years earlier than current practice.

Keywords: nano-biosensor; obesity-related cancer; inflammatory cytokines; extracellular vesicles; point-of-care diagnostics

INTRODUCTION

The mechanistic bridge between obesity and cancer is constructed from persistent immune activation, endocrine imbalance, and metabolic rewiring[1]. Hypertrophic adipocytes develop hypoxia and endoplasmic-reticulum stress, recruiting macrophages and neutrophils that amplify cytokine cascades and remodel tissue architecture. Insulin resistance elevates insulin and IGF-1 tone; adipokine balance shifts toward leptin dominance and adiponectin deficiency; lipotoxic species such as ceramides accumulate[2-4]. Together these processes sculpt a pre-malignant niche characterized by fibroinflammation, aberrant angiogenesis, and immune suppression. Biomarkers that report on these axes emerge in blood, interstitial fluid, saliva, urine, and sweat: interleukins and TNF family members; leptin and adiponectin; chemokines and prostaglandins; metabolic intermediates such as lactate and specific lipid species; extracellular vesicles carrying microRNAs and proteins; and tumor-derived DNA fragments with characteristic methylation or mutation patterns[5, 6]. The clinical challenge is to measure them early and in combination, when intervention can reverse risk or detect cancer at curable stages.

Conventional immunoassays and mass spectrometry define gold standards for accuracy, yet they are resource intensive, slow, and often optimized for single analytes[6]. They also struggle when analyte concentrations fall into the low pg mL^{-1} to fg mL^{-1} range or when matrices are hostile because of dyslipidemia, glycation, and elevated acute-phase proteins common in obesity. Nanotechnology reframes the measurement problem[7]. By shrinking transducer dimensions to the scale of recognition events, nano-biosensors translate binding, catalytic, or structural changes into large electrical, optical, or mechanical signals. Graphene, carbon nanotubes, and semiconducting nanowires offer ballistic or near-ballistic charge transport so that single binding events shift conductance measurably[8-11]. Noble-metal nanostructures sustain localized surface plasmon resonances and intense near-fields that amplify refractive index changes or Raman scattering from a handful of molecules. Solid-state nanopores and plasmonic nanoholes discriminate nucleic acids, vesicles, and lipoproteins by size, charge, and optical signatures[12-15]. Micro- and nanofluidics concentrate targets from tiny volumes, sort vesicles by size or affinity, and deliver them to sensing sites with minimal loss.

Early detection at the obesity cancer interface is a multiplex problem, not a single-analyte one. A convincing signature might pair a rise in IL-6 and leptin with a drop in adiponectin, a surge in specific ceramides, and an extracellular-vesicle miRNA pattern—all changing subtly before imaging detects a lesion[16]. Nano-biosensors are increasingly engineered to read such panels through spatial arrays, barcoded nanoparticles, ratiometric optical outputs, and multi-electrode impedance maps[17]. The remaining obstacles are translation hurdles: calibrating sensors in dyslipidemic plasma, stabilizing antifouling layers without sacrificing sensitivity, standardizing pre-analytics, and embedding analytics that separate signal from patient-specific background. The subsequent sections delineate the biomarker landscape most relevant to obesity-linked oncogenesis, the principal nano-biosensing modalities that can detect them, strategies for extracellular-vesicle and nucleic-acid capture, routes to minimally invasive and wearable monitoring, and the validation ecosystem needed to convert prototypes into practice.

2 The Biomarker Landscape Connecting Obesity and Cancer

Inflammatory cytokines such as IL-6, TNF- α , IL-1 β , and chemokines like CCL2 rise with adiposity and predict cancer risk and progression; they drive STAT3, NF- κ B, and AP-1 programs in epithelial and stromal cells[18–20]. Adipokines reflect endocrine tone: leptin elevations correlate with angiogenesis and epithelial–mesenchymal transition, while adiponectin reductions remove AMPK-mediated brakes on mTORC1, oxidative stress, and inflammation. Oncometabolites form a second stratum[18]. Persistent hypoxia and high glycolytic flux increase lactate, acidify interstitium, and shape immune suppression; lipidomic signatures particularly select ceramides and acylcarnitines mirror lipotoxic stress and link to hepatocarcinogenesis and colorectal neoplasia. Extracellular vesicles released by adipocytes, immune cells, and nascent tumors traffic miRNAs, proteins, and lipids that instruct angiogenesis, matrix remodeling, and immune evasion; vesicle counts and cargo composition shift months before imaging findings[21]. Circulating tumor DNA adds mutation and methylation layers; in obesity, total cell-free DNA may increase through adipose inflammation, demanding sensors with strong discrimination of tumor-specific features[21].

From a sensing perspective, this palette demands single-digit pg mL⁻¹ limits of detection for cytokines, sub-nanomolar for adipokines, micromolar precision for lactate in complex fluids, and vesicle and DNA analytics at the 10⁴–10⁶ particles μ L⁻¹ and <0.1% variant allele frequency levels[22]. It also requires antifouling strategies that preserve specificity despite abundant lipoproteins, albumin, and glycation products. The convergence of recognition chemistry, nanostructured transducers, and microfluidic preprocessing is essential to meet these constraints without sacrificing speed or affordability.

3 Electrochemical and Field-Effect Nano-Transducers for Cytokines and Adipokines

Electrochemical platforms translate molecular recognition into current, potential, or impedance shifts with sub-minute time constants. Nanostructuring the electrode with graphene, carbon nanotubes, gold nanoprisms, or porous platinum increases active area and electron-transfer rates, lowering noise and pushing limits of detection toward the single-molecule regime in optimized conditions. For IL-6, TNF- α , leptin, and adiponectin, antibody- or aptamer-functionalized microarrays on such nanostructured electrodes enable simultaneous, label-free quantification across clinically relevant ranges[12–14, 23]. Redox reporters tethered to aptamers supply ratiometric readouts that self-normalize for matrix variability. Non-enzymatic lactate sensing with nanoporous platinum or nickel nanostructures avoids enzyme drift and supports continuous monitoring; enzyme-based designs with lactate oxidase gain selectivity but must mitigate peroxide-induced electrode aging, a problem reduced by catalytic nanocoatings.

Field-effect transistor (FET) biosensors exploit the gate-modulating effect of charged analytes binding within the Debye length of a semiconductor channel. Graphene and molybdenum disulfide FETs functionalized with cytokine or adipokine aptamers report femtomolar binding as real-time conductance shifts[8, 24–26]. Debye screening in high-ionic-strength samples is managed by nanogap architectures, hydrogel spacers, or on-chip desalting. For panels, crossbar arrays of FETs with orthogonal capture chemistries deliver multi-analyte fingerprints in a single drop of plasma or saliva. To survive dyslipidemia, these devices depend on zwitterionic brushes, poly(ethylene glycol) alternatives such as poly(2-oxazoline), or peptide-based antifouling monolayers that resist non-specific adsorption while maintaining aptamer mobility.

4. Plasmonic, SERS, and Quantum-Dot Strategies for Label-Free and Ratiometric Readouts

Localized surface plasmon resonance (LSPR) sensors built from gold nanorods, nanoholes, and nanoprisms detect refractive-index changes induced by biomolecular binding with picogram-per-milliliter sensitivity[26–28]. Their optical nature suits transparent microfluidic integration and supports real-time kinetic profiling to distinguish high- from low-affinity interactions, useful when cytokine family cross-reactivity threatens specificity. Surface-enhanced Raman scattering (SERS) leverages electromagnetic hot spots between metal nanostructures to magnify molecular vibrational signatures, enabling multiplex detection via spectral barcodes. Sandwich assays that pair capture antibodies or aptamers with SERS-encoded nanoparticles read multiple cytokines and adipokines in a single spectrum, minimizing sample volume and eliminating wash steps when coupled with magnetic pre-concentration[29–31].

Quantum-dot immunoassays extend dynamic range through photostable, spectrally narrow emissions. Ratiometric quantum-dot FRET designs place donor and acceptor dots across a binding interface so that target engagement produces predictable energy-transfer changes resistant to intensity fluctuations[32]. In saliva or

sweat where analyte levels are lower and ionic backgrounds fluctuate, ratiometry enhances robustness. Optical sensors also pair naturally with smartphone cameras or compact readers, enabling decentralized screening in metabolic clinics without compromising analytical performance.

5. Extracellular Vesicle and Circulating Nucleic-Acid Sensing with Nanopores and Microfluidics

Extracellular vesicles concentrate the signals of the obesity–cancer axis but are heterogeneous and easily confounded by lipoproteins[33–35]. Affinity-based nanointerfaces using anti-tetraspanin antibodies, adipocyte- or tumor-selective lectins, or engineered aptamers enrich target vesicles on nanoporous gold or silica, while acoustofluidic and deterministic lateral displacement modules fractionate by size and deformability[30, 36, 37]. On-chip lysis followed by nano-sensing of miRNAs with catalytic hairpin amplification on graphene electrodes or SERS-encoded probes permits detection without PCR[24, 38, 39]. For circulating tumor DNA, solid-state nanopores distinguish single-molecule events by ionic current blockades, and plasmonic nanohole arrays boost optical sensitivity for methylation-specific recognition. CRISPR-Cas trans-cleavage readouts on nanostructured electrodes or quantum dots add sequence specificity at low variant allele fractions, enabling the separation of tumor-derived DNA from obesity-related background increases in cell-free DNA.

Critical to translation is pre-analytics. Microfluidic cartridges that standardize dilution, lipid clearing, and anticoagulant exposure reduce inter-site variability[38]. On-cartridge calibration using internal nanoparticle standards corrects for matrix-dependent signal drift. Such standardization is particularly important in populations with high triglycerides, altered lipoprotein profiles, and variable acute-phase reactants that otherwise erode assay specificity.

6 Toward Longitudinal Monitoring: Wearables, Microneedles, Data Fusion, and Translation

Because obesity and its inflammatory outputs evolve over months to years, longitudinal tracking may reveal risk transitions invisible to single draws[40]. Microneedle arrays accessing interstitial fluid allow painless, repeatable sampling near subcutaneous adipose where cytokine and adipokine dynamics first shift. Embedding electrochemical or LSPR nanosensors at the microneedle tip yields continuous or on-demand readouts, supported by capillary-driven microfluidics and passive antifouling coatings[40]. Sweat and saliva provide additional, low-barrier matrices; while absolute concentrations differ from plasma, machine-learning models can learn transformation functions from paired samples to infer plasma-equivalent values. Multimodal fusion that combines cytokines, adipokines, lactate, selected lipids, and vesicle markers improves predictive value and reduces false positives.

Clinical adoption rests on calibration, quality-by-design manufacturing, and regulatory science. Sensors must be calibrated against reference methods across BMI strata to derive obesity-adjusted decision thresholds[41]. Antifouling and capture chemistries require batch-to-batch reproducibility under stress tests that mimic dyslipidemic plasma and high fibrinogen states. Analytical validity precedes clinical validity: prospective studies should demonstrate that nano-biosensor signatures anticipate imaging or biopsy findings and that intervening on elevated signatures modifies outcomes[41]. Data security and privacy are non-negotiable for wearable deployments, and algorithms must guard against bias by ensuring training sets represent diverse body compositions and comorbidity profiles.

CONCLUSION

Nano-biosensing has matured from proof-of-concept physics to platform candidates capable of reading the complex biology that links obesity to cancer. Electrochemical and field-effect devices convert scarce cytokines and adipokines into robust electrical signals in microliter samples. Plasmonic, SERS, and quantum-dot optics provide label-free or ratiometric measurements suitable for multiplex panels and decentralized settings. Nanopore and microfluidic systems interrogate extracellular vesicles and tumor-derived nucleic acids with the specificity needed to stand out in inflamed, lipid-rich backgrounds. When coupled to antifouling surface science, rigorous pre-analytics, and longitudinal, minimally invasive sampling, these technologies can detect actionable shifts in the obesity-cancer axis long before structural disease is apparent. The remaining work—BMI-aware calibration, manufacturing discipline, and prospective validation—will convert these nanoscale advantages into earlier diagnoses, better risk stratification, and timelier interventions for patients living with obesity.

REFERENCES

1. Kompella, P., Vasquez, K.M.: Obesity and Cancer: a mechanistic overview of metabolic changes in obesity that impact genetic instability. *Mol. Carcinog.* 58, 1531–1550 (2019). <https://doi.org/10.1002/mc.23048>
2. Abdillahi, A.M., Yun, J.W.: Capsaicin induces ATP-dependent thermogenesis via the activation of TRPV1/ β 3-AR/ α 1-AR in 3T3-L1 adipocytes and mouse model. *Arch. Biochem. Biophys.* 755, 109975 (2024). <https://doi.org/10.1016/j.abb.2024.109975>
3. Chang, J.S.: Recent insights into the molecular mechanisms of simultaneous fatty acid oxidation and synthesis in brown adipocytes. *Front. Endocrinol.* 14, (2023). <https://doi.org/10.3389/fendo.2023.1106544>
4. Alum, E.U., Obasi, D.C., Abba, J.N., Aniketete, U.C., Okoroh, P.N., Akwari, A.Ak.: Evolving Paradigms in Nutrition Therapy for Diabetes: From Carbohydrate Counting to Precision Diets. *Obes. Med.* 100622 (2025). <https://doi.org/10.1016/j.obmed.2025.100622>
5. Skapinker, E., Aucoin, E.B., Kombargi, H.L., Yaish, A.M., Li, Y., Baghaie, L., Szewczuk, M.R.: Contemporaneous Inflammatory, Angiogenic, Fibrogenic, and Angiostatic Cytokine Profiles of the Time-

- to-Tumor Development by Cancer Cells to Orchestrate Tumor Neovascularization, Progression, and Metastasis. *Cells*. 13, 1739 (2024). <https://doi.org/10.3390/cells13201739>
6. Bocian-Jastrzębska, A., Malczewska-Herman, A., Kos-Kudła, B.: Role of Leptin and Adiponectin in Carcinogenesis. *Cancers*. 15, 4250 (2023). <https://doi.org/10.3390/cancers15174250>
 7. Vekic, J., Vujcic, S., Bufan, B., Bojanin, D., Al-Hashmi, K., Al-Rasadi, K., Stoian, A.P., Zeljkovic, A., Rizzo, M.: The Role of Advanced Glycation End Products on Dyslipidemia. *Metabolites*. 13, 77 (2023). <https://doi.org/10.3390/metabo13010077>
 8. Şak, B., Sousa, H.B.A., Prior, J.A.V.: Carbon Nanomaterial-Based Electrochemical Biosensors for Alzheimer's Disease Biomarkers: Progress, Challenges, and Future Perspectives. *Biosensors*. 15, 684 (2025). <https://doi.org/10.3390/bios15100684>
 9. Al Tahan, M.A., Al-Khattawi, A., Russell, C.: Oral peptide delivery Systems: Synergistic approaches using polymers, lipids, Nanotechnology, and needle-based carriers. *J. Drug Deliv. Sci. Technol.* 112, 107205 (2025). <https://doi.org/10.1016/j.jddst.2025.107205>
 10. Azmi, N.A.N., Elgharbawy, A.A.M.: Advances in Medical Applications: The Quest of Green Nanomaterials. In: Shanker, U., Hussain, C.M., and Rani, M. (eds.) *Handbook of Green and Sustainable Nanotechnology: Fundamentals, Developments and Applications*. pp. 1889–1909. Springer International Publishing, Cham (2023)
 11. Bishoyi, A.K., Nouri, S., Hussen, A., Bayani, A., Khaksari, M.N., Soleimani Samarkhazan, H.: Nanotechnology in leukemia therapy: revolutionizing targeted drug delivery and immune modulation. *Clin. Exp. Med.* 25, 166 (2025). <https://doi.org/10.1007/s10238-025-01686-z>
 12. Behzadifar, S., Barras, A., Plaisance, V., Pawlowski, V., Szunerits, S., Abderrahmani, A., Boukherroub, R.: Polymer-Based Nanostructures for Pancreatic Beta-Cell Imaging and Non-Invasive Treatment of Diabetes. *Pharmaceutics*. 15, 1215 (2023). <https://doi.org/10.3390/pharmaceutics15041215>
 13. Mhlanga, N., Mphuthi, N., Van der Walt, H., Nyembe, S., Mokhena, T., Sikhwivhilu, L.: Nanostructures and nanoparticles as medical diagnostic imaging contrast agents: A review. *Mater. Today Chem.* 40, 102233 (2024). <https://doi.org/10.1016/j.mtchem.2024.102233>
 14. Sharma, S., Kumari, R., Varshney, S.K., Lahiri, B.: Optical biosensing with electromagnetic nanostructures. *Rev. Phys.* 5, 100044 (2020). <https://doi.org/10.1016/j.revip.2020.100044>
 15. Wang, T., Liu, Y., Wu, Q., Lou, B., Liu, Z.: DNA nanostructures for stimuli-responsive drug delivery. *Smart Mater. Med.* 3, 66–84 (2022). <https://doi.org/10.1016/j.smaim.2021.12.003>
 16. Salemi, R., Sergi, V., Basile, M.S., Bravaccini, S., Frittitta, L., Graziano, A.C.E., Filippello, A., Malaguarnera, R., De Francesco, E.M.: Microenvironmental determinants of cancer progression during obesity: emerging evidence and novel perspectives. *J. Transl. Med.* 23, 995 (2025). <https://doi.org/10.1186/s12967-025-06970-w>
 17. Nalabolu, M.R., Palasamudram, K., Jamil, K.: Adiponectin and Leptin Molecular Actions and Clinical Significance in Breast Cancer. *Int. J. Hematol.-Oncol. Stem Cell Res.* 8, 31–40 (2014)
 18. Uti, D.E., Atangwho, I.J., Omang, W.A., Alum, E.U., Obeten, U.N., Udeozor, P.A., Agada, S.A., Bawa, I., Ogbu, C.O.: Cytokines as key players in obesity low grade inflammation and related complications. *Obes. Med.* 54, 100585 (2025). <https://doi.org/10.1016/j.obmed.2025.100585>
 19. Divella, R., De Luca, R., Abbate, I., Naglieri, E., Daniele, A.: Obesity and cancer: the role of adipose tissue and adipo-cytokines-induced chronic inflammation. *J. Cancer*. 7, 2346–2359 (2016). <https://doi.org/10.7150/jca.16884>
 20. Alum, E.U., Uti, D.E., Ugwu, O.P.-C., Alum, B.N., Edeh, F.O., Ainebyoona, C.: Unveiling the microbial orchestra: exploring the role of microbiota in cancer development and treatment. *Discov. Oncol.* 16, 646 (2025). <https://doi.org/10.1007/s12672-025-02352-2>
 21. Singh, L., Nair, L., Kumar, D., Arora, M.K., Bajaj, S., Gadewar, M., Mishra, S.S., Rath, S.K., Dubey, A.K., Kaithwas, G., Choudhary, M., Singh, M.: Hypoxia induced lactate acidosis modulates tumor microenvironment and lipid reprogramming to sustain the cancer cell survival. *Front. Oncol.* 13, 1034205 (2023). <https://doi.org/10.3389/fonc.2023.1034205>
 22. Rose, G.L., Farley, M.J., Flemming, N.B., Skinner, T.L., Schaumberg, M.A.: Between-day reliability of cytokines and adipokines for application in research and practice. *Front. Physiol.* 13, 967169 (2022). <https://doi.org/10.3389/fphys.2022.967169>
 23. Elblová, P., Anthi, J., Liu, M., Lunova, M., Jirsa, M., Stephanopoulos, N., Lunov, O.: DNA Nanostructures for Rational Regulation of Cellular Organelles. *JACS Au*. 5, 1591–1616 (2025). <https://doi.org/10.1021/jacsau.5c00117>
 24. Alghannam, F., Alayed, M., Alfihed, S., Sakr, M.A., Almutairi, D., Alshamrani, N., Al Fayez, N.: Recent Progress in PDMS-Based Microfluidics Toward Integrated Organ-on-a-Chip Biosensors and Personalized Medicine. *Biosensors*. 15, 76 (2025). <https://doi.org/10.3390/bios15020076>
 25. Animashaun, C., Lahcen, A.A., Slaughter, G.: Gold Nanoparticle-Enhanced Molecularly Imprinted Polymer Electrode for Non-Enzymatic Lactate Sensing. *Biosensors*. 15, 384 (2025). <https://doi.org/10.3390/bios15060384>

26. Beeram, R., Vepa, K.R., Soma, V.R.: Recent Trends in SERS-Based Plasmonic Sensors for Disease Diagnostics, Biomolecules Detection, and Machine Learning Techniques. *Biosensors*. 13, 328 (2023). <https://doi.org/10.3390/bios13030328>
27. Demishkevich, E., Zyubin, A., Seteikin, A., Samusev, I., Park, I., Hwangbo, C.K., Choi, E.H., Lee, G.J.: Synthesis Methods and Optical Sensing Applications of Plasmonic Metal Nanoparticles Made from Rhodium, Platinum, Gold, or Silver. *Materials*. 16, 3342 (2023). <https://doi.org/10.3390/ma16093342>
28. Hlapisi, N., Songca, S.P., Ajibade, P.A.: Capped Plasmonic Gold and Silver Nanoparticles with Porphyrins for Potential Use as Anticancer Agents—A Review. *Pharmaceutics*. 16, 1268 (2024). <https://doi.org/10.3390/pharmaceutics16101268>
29. Cennamo, N., Pesavento, M., Arcadio, F., Morrone, B., Seggio, M., Zeni, L.: Plasmonic sensor combined with a microcuvette device for monitoring molecule binding processes at ultra-low concentrations. *Sens. Actuators B Chem.* 416, 136050 (2024). <https://doi.org/10.1016/j.snb.2024.136050>
30. Hang, Y., Wang, A., Wu, N.: Plasmonic silver and gold nanoparticles: shape- and structure-modulated plasmonic functionality for point-of-care sensing, bio-imaging and medical therapy. *Chem. Soc. Rev.* 53, 2932–2971 (2024). <https://doi.org/10.1039/d3cs00793f>
31. Shoukat, C.A., Tariq, M., Aqib, R.M., Tajwar, M.A., Iqbal, R.: Plasmonic ELISA for Biomarker Detection: A Review of Mechanisms, Functionalization Strategies, and Emerging Modalities. *ACS Appl. Bio Mater.* (2025). <https://doi.org/10.1021/acsabm.5c00738>
32. Kalvaityte, U., Bagdonas, E., Kirdaite, G., Kausaite-Minkstimiene, A., Uzieliene, I., Ramanaviciene, A., Popov, A., Butkiene, G., Karabanovas, V., Denkovskij, J., Mobasheri, A., Bernotiene, E.: Development of a Sensitive Quantum Dot-Linked Immunoassay for the Multiplex Detection of Biochemical Markers in a Microvolumetric Format. *Int. J. Nanomedicine*. 20, 1717–1729 (2025). <https://doi.org/10.2147/IJN.S477118>
33. Adlerz, K., Patel, D., Rowley, J., Ng, K., Ahsan, T.: Strategies for scalable manufacturing and translation of MSC-derived extracellular vesicles. *Stem Cell Res.* 48, 101978 (2020). <https://doi.org/10.1016/j.scr.2020.101978>
34. Clement, E., Lazar, I., Attané, C., Carrié, L., Dauvillier, S., Ducoux-Petit, M., Esteve, D., Menneteau, T., Moutahir, M., Le Gonidec, S., Dalle, S., Valet, P., Burlet-Schiltz, O., Muller, C., Nieto, L.: Adipocyte extracellular vesicles carry enzymes and fatty acids that stimulate mitochondrial metabolism and remodeling in tumor cells. *EMBO J.* 39, e102525 (2020). <https://doi.org/10.15252/embj.2019102525>
35. Hánělová, K., Raudenská, M., Masařík, M., Balvan, J.: Protein cargo in extracellular vesicles as the key mediator in the progression of cancer. *Cell Commun. Signal.* 22, 25 (2024). <https://doi.org/10.1186/s12964-023-01408-6>
36. Mangadla, J.D., Wang, X., McCleese, C., Escamilla, M., Ramamurthy, G., Wang, Z., Govande, M., Basilion, J.P., Burda, C.: Prostate-Specific Membrane Antigen Targeted Gold Nanoparticles for Theranostics of Prostate Cancer. *ACS Nano*. 12, 3714–3725 (2018). <https://doi.org/10.1021/acsnano.8b00940>
37. Jeong, S., Park, S.-H., Lee, S., Cho, H., Lee, K., Ju, B.K., Lee, Y.J., Lee, S.H.: Electrochemical biosensor based on gold nanoparticles/laser induced graphene for diagnosis of Parkinson's disease by detecting phosphorylated α -synuclein in human blood. *Chem. Eng. J.* 509, 161329 (2025). <https://doi.org/10.1016/j.cej.2025.161329>
38. Cetinkaya, A., Kaya, S.I., Ozkan, S.A.: A review of point-of-care (POC) and lab-on-chip (LOC) approaches in molecularly imprinted polymer-based electrochemical sensors for biomedical applications. *Anal. Chim. Acta.* 1357, 344080 (2025). <https://doi.org/10.1016/j.aca.2025.344080>
39. Sampaio, A.R., Maia, R.F., Ciardulli, M.C., Santos, H.A., Sarmiento, B.: Organ-on-chip platforms for nanoparticle toxicity and efficacy assessment: Advancing beyond traditional in vitro and in vivo models. *Mater. Today Bio.* 33, 102053 (2025). <https://doi.org/10.1016/j.mtbio.2025.102053>
40. Zhao, X., Sun, X., Wu, F., Huang, K., Jing, X., Lv, M., Zhu, J., Li, J., Jing, F., Mao, Y., Ye, D.: Longitudinal patterns of obesity index changes and risk of incident rheumatoid arthritis: evidence from a population-based cohort. *Arthritis Res. Ther.* 27, 186 (2025). <https://doi.org/10.1186/s13075-025-03655-z>
41. Kaplan, L.M., Apovian, C.M., Ard, J.D., Allison, D.B., Aronne, L.J., Batterham, R.L., Busetto, L., Dicker, D., Horn, D.B., Kelly, A.S., Mechanick, J.I., Purnell, J.Q., Ramos-Salas, X.: Assessing the state of obesity care: Quality, access, guidelines, and standards. *Obes. Sci. Pract.* 10, e765 (2024). <https://doi.org/10.1002/osp4.765>

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